# Some biochemical effects of *Aloe vera* leaves on tissues in normal mice

# M. G. Matti, S. A. Al-Ameen and S. H. Rashed

Department of Chemistry, College of Science, University of Mosul, Mosul, Iraq

(Received March 2, 2009; Accepted January 17, 2010)

#### **Abstract**

This research was carried out mainly to investigate the effects of the aqueous extract and proteinous fractions of *Aloe vera* leaves on cholesterol, acetycholinesterase in brain, glycogen, glutathione in liver and malonaldehyde levels in heart in normal male albino mice. The antioxidant properties and inhibition of acetylcholine-sterase in tissue were detected. Intraperitoneal administration of *Aloe vera* extract in concentration of 400 mg/kg significantly decreased the levels of AchE in brain by (-88.27%) and glutathione content in liver by (-35.48%), and increased the levels of glycogen in liver and malonaldehyde in heart by (22.60%, 85.50%) respectively. At a concentration of 300 mg/kg *Aloe vera* extract significantly increased the level of cholesterol in brain by (24.39%). These results clearly show the antioxidant property of the extract of *Aloe vera* leaves.

**Keywords:** Aloe vera; Aqueous extract; Brain; Liver. Available online at <a href="http://www.vetmedmosul.org/ijvs">http://www.vetmedmosul.org/ijvs</a>

· ( / ) % , - % , - %

# Introduction

Aloe vera is the gift of nature, it is a miracle been used medicinally for centuries. It belongs to the family (Liliaceae) (1). Botanically known as Aloe barbadensis, Mill (2).

The medicinal properties of various parts (Leaf, Aloe gel) are rich in minerals (magnesium, calcium, chromium, copper, iron,...etc), enzymes, amino acids, vitamins as antioxidants (3). *Aloe vera* plays an important role in many

medical properties as a traditional medicine in treatment of many physiological disease and conditions such as Insomnia, Burns, Ulcerative colitis, Psoriasis, wound healing (4). Despite the fact that more than 200 plants are used around the world in the empirical control of antioxidant effect, most of them have not been pharmacologically and chemically investigated (5). Oxidative stress in cells and tissues results from the increased generation of reactive oxygen species and /or from decrease in antioxidant defense potential (6). The

antioxidant defense system represents a complex network with interactions, synergy and specific tasks for a given antioxidant (7). The plant contains substantial amounts of antioxidant including vitamin E, vitamin C, flavenoids, carotenoids and tannins (8).

The present investigation was carried out to study the effect of the aqueous extract and proteinous fractions of *aloe vera* on tissues glycogen, glutathione, malonaldehyde, acetylcholinesterase, cholesterol in experimental mice.

#### Material and methods

#### **Plant**

The plant was collected from the nursery of the College of Agriculture and Forests, University of Mosul and was classified according to plant taxonomy or plant classification references related to medicinal plants. Scientific name = (*Aloe vera*) also known (Aloe barbadensis) (9). Common name = True Aloe or Barbados Aloe. Family = Liliaceae (10).

### Preparation of aqueous extract

The cold extract of the plant was prepared by grindingfresh leaves of *Aloe vera* for 10 minutes using a blender, then frozen in a deep freezer, sufficient amount of distilled water was added and the crude homogenate was stirred for additional one hour then filtered through glass wool. The mixture was centrifuged for 15 minutes at 4000xg. The filtrate Concentrated to one third volume by lyophilizer then kept until investigation.

## **Preparation of proteinous fractions**

Proteinous precipitate was separated by full saturation of ammonium sulphate (75%) (11). Gradual addition of ammonium sulphate at 4°C for 60 minutes accompanied with mixing by electrical mixer. The mixture was left at 0 °C for 24 hours and then the proteinous precipitate was isolated by centrifugation for 20 minutes at 6000xg at 4 °C. The precipitate was dried by lyophilizer then kept for gel filtration chromatography. Total protein concentration in each step was determined by modified –lowery method (12).

#### Gel filtration chromatography

Total protein was fractionated by gel filtration chromatography using sephadex G-100 in column (2×88 cm). Distilled water was used as an eluent solution. The same technique was used to determine the comparative molecular weights of proteinous fractions (13).

The molecular weight of each proteinous compound (comparative) was obtained from its elution volume under the same conditions of known molecular weights such as (Blue dextran, Bovine serum albumin, Eggs albumin, Trypsin, Insulin, Tryptophan).

#### Animals

Forty eight male albino mice were obtained from the animal house of College of Education, University of Mosul, at 2-2.5 months of age, weighing 22.5±2 g. They were housed in polypropylene cages under standard conditions at temperature 27 °C and had free access to food and water.

The mice were divided randomly into twelve groups (4mice/group). The first group received distilled water and served as control. Groups (two, three and four) were administered orally with gavages needle in increasing concentration (300, 400 and 500) mg/kg of body weight of crude extract of the plant (14). Groups (five and six) were administered orally with dose of (40 mg/kg) of body weight of proteinous peak I, II respectively after fasting for (16) hours (15). Another set of animals (group eight, nine and ten) were administered intraperitoneally in increasing concentration (300, 400 and 500) mg/kg of body weight of crude extract. Group (eleven and twelve) were administered intraperitoneally with dose of (40mg/kg) of body weight of proteinous peak I, II respectively after fasting (16) hours. While the group seven was kept as control and administered intraperitoneally with distilled water. After two hours of administration, blood samples were collected by orbital sinus puncture technique after ether anesthesia (16). The liver, heart and brain of these mice were immediately removed after sacrification washed with ice-cold saline and kept frozen (-10 C) until glycogen, glutathione, malonaldehyde, cholesterol and acetyl cholinesterase contents were estimated.

### Parameters assayed

UV-visible CECIL CE 1021 single beam spectrophotometer were used to determine all parameters. Serum was separated by centrifugation at (4000xg). Serum and brain cholesterol was determined using (BLOLABO SA. Maizy, France) (17). Glycogen content in liver was determined by Colorimetric method (18). Glutathione content in liver was determined and quantitated by modified Ellman method (19). Malonaldehyde content in heart was determined by spectrophotometric method (20). Acetyl cholinesterase assayed using spectrophotometric modification method after slight (21)acetylthiocholine iodide (AschI) as a substrate.

# Statistical analysis

The statistical methods used to analize data and estimate mean and standard error. Student's-T-test was used to compare between control and experimental mice was at significance of level (P<0.05) (22).

### Results

Mean values of blood cholesterol, brain cholesterol and acetylcholinesterase, liver glycogen and glutathione, heart malonaldehyde for control and different doses of *Aloe vera* (oral administered) for normal mice shown in (Table 1). Mean values of blood cholesterol and other parameters in

tissues for control and different doses of *Aloe vera* (Intraperitoneal administered) for normal mice shown in (Table 2).

Table 1: Mean blood serum cholesterol, liver glycogen and glutathione content, heart malonaldehyde content, brain cholesterol, acetyl cholinesterase levels in mg/dl±SE after oral administration of different doses of crude extract and proteinous materials of *Aloe vera* in normal fasted mice.

Group No.	Treatment	Glutathione content liver nmol/g±SE	Change %	Malonal- dehyde heart μmol/L ±SE	Change %	Glycogen content liver mg/g±SE	Change %	AchE brain μmol/min/ml ±SE	Change %	Cholesterol brain mg/dl±SE	Change %	Serum Cholesterol level mg/dl ±SE	Change %
		33.19		14.04		2.11		0.37		152.02		173.69	
1	Control	±		±		±		±		±		±	
		1.14		0.38		5.7		0.14		8.51		6.00	
	Crude	35.70		18.86		2.12		0.27		181.16		200.23	
2	extract	±	5.66	±	34.35	±	0.47	±	-27.37	±	19.16	±	15.28
	300mg/10ml	0.97		1.22		6.5		0.11				5.84	
	Crude	31.19	- 0.	21.60	40.4=	2.29	0.50	0.53	40.00	168.11	40.50	209.88	•••
3	extract	±	-6.02	±	49.67	±	8.53	±	42.28	±	10.58	±	20.84
	400mg/10ml	0.82		0.88		9.4		0.16		7.27		16.76	
	Crude	32.33	2.50	19.46	20.70	2.22	5.01	0.19	46.24	170.0	11.00	131.10	24.5
4	extract	±	-2.59	±	38.70	±	5.21	±	46.34	±	11.82	±	-24.5
	500mg/10ml	0.83		0.94		9.5		0.12		4.95		10.43	
_	Proteinous	34.22	2 10	20.27	44.20	2.05	2.04	0.30	12.60	156.55	2.07	199.10	14.62
5	Frc.I	± 0.93	3.10	± 1.04	44.39	± 3.3	-2.84	± 0.02	12.68	± 7.19	2.97	± 7.69	14.63
	40mg/10 ml Proteinous												
6	Fre.II	33.89 ±	2.10	12.64 ±	-9.85	2.25 ±	6.63	0.33 ±	11.65	172.71 ±	13.61	211.36 ±	21.69
U	40mg/10 ml	± 1.01	2.10	1.23	-9.03	3.3	0.03	± 0.1	11.03	7.08	13.01	± 6.95	21.09
	Tomg/ to illi	1.01		1.43		ر. د		0.1		7.00		0.33	

The values are mean  $\pm$  SE of 4 mice each group.

#### **Discussion**

In the present study, the activity of cholinergic neurotransmission following previous administration of increasing doses of crude extract and proteinous fractions I, II of *aloe vera* plant were investigated by assaying brain AchE activity. Results depicted in Tables 1 and 2 showed (group 9) inhibition of brain cholinesterase activity compared to the values of control group. The percentage of inhibition suppression of activity of AchE was (-88.27%). The inhibitory effect of curde extract (400 mg/kg) of *aloe vera* may be due to inhibition at the anionic site of AchE (23). On the other hand, many reports suggest that such inhibition is due to a decrease in the internal microviscosity of phospholipids leading to change in the fluidity of microsomal membranes of the brain (24).

The results in (Tables 1 and 2) also indicate that there is significant increase the brain cholesterol levels after intraperitoneal administration of curde extract (300 mg/kg) by (24.39%) in normal mice. The high fiber content of *Aloe vera* had been shown to exert beneficial effects on activation of the hydroxy methyl glutaryl-CoA reductase (HMG-CoA reductase), this enzyme is necessary for cholesterol biosynthesis (25).

The mean values of heart malonaldehyde (MAD) in control and *Aloe vera* extracts treated normal mice are shown in tables (1 and 2). The results illustrate a significant increase of MAD in group 9 at (P<0.05) by intraperitoneal administration of curde extract (400 mg/kg) by (85.5%) in normal mice compared to control group.

The increase of MAD may be due to increase of lipid peroxidation, so the higher level of glucose in blood due to

elevation of proliferative free radical as a result of, this due to increased lipid peroxidation (26).

Mice treated with *Aloe vera* extract at a dose of 400 mg/kg showed significant increase in liver glycogen contents at a level of 22.6% compared with control value, this may be to that the extract can activate glycogen synthytase leading to synthesize of glycogen and inhibition of glycogenolysis leading in turn to gluconeogensis or release insulin from  $\beta$ -cells in pancrease (27).

Glutathione is tripeptide normally present at high concentrations intracellularly, and constitutes the major

reducing capacity of the cytoplasm. Glutathione is known to protect the cellular system against toxic effect of lipid peroxidation (28). The extract of *Aloe Vera* at 400 mg/kg orally administered indicate significant decrease of GsH level by (35.48%) compared to control value. This decrease may be due to decline in Glutathione peroxidase (GPx) activity (29) or activity of GsH to prevent oxidation in oxidative stress through elimination of free radical so the level of GsH is decreased (30).

Table 2: Mean blood serum cholesterol, liver glycogen and glutathione content, heart malonaldehyde content, brain cholesterol, acetyl cholinesterase levels in mg/dl±SE after intraperitoneal administration of different doses of crude extract and proteinous materials of *Aloe vera* in normal mice.

Group No.	Treatment	Glutathione content liver nmol/g±SE	Change %	Malonal- dehyde heart μmol/L ±SE	Change %	Glycogen content liver mg/g±SE	Change %	AchE brain μmol/min/ml ±SE	Change %	Cholesterol brain mg/dl±SE	Change %	Serm Cholesterol level mg/dl ±SE	Change %
_	a	35.87		24.71		2.87		0.32		148.04		173.68	
7	Control	±		±		±		±		±		±	
	Consider	0.82		0.62		11.2		0.12		11.96		13.30	
0	Crude	32.75	0.60	31.05	25.65	2.65	7.00	0.26	17.20	*184.1	24.20	196.64	12.21
8	extract	±	-8.69	±	25.65	±	-7.66	±	-17.28	±	24.39	±	13.21
	300mg/10ml	0.67		1.44		6.9		0.15		6.71		8.01	
0	Crude	*23.1	25.4	*45.8	05.50	*3.5	22.6	*0.04	00.27	115.58	21.0	203.66	17.26
9	extract	±	-35.4	±	85.50	±	22.6	±	-88.27	±	-21.9	±	17.26
	400mg/10ml	0.91		2.23		8.6		0.03		7.32		17.89	
10	Crude	33.27	7.24	33.55	25 77	3.04	5.00	0.22	20.02	170.95	15 47	*244.4	40.72
10	extract	± 0.48	-7.24	± 2.45	35.77	± 2.9	5.92	± 0.12	-29.93	± 4.76	15.47	± 12.39	40.72
	500mg/10ml Proteinous	28.67		22.29		2.93		0.12		162.22		12.39	
11	Frc.I	28.07 ±	-20.0	±	-9.79	2.93 ±	2.09	0.22 ±	-31.48	±	9.57	±	-19.9
11	40mg/10 ml	0.85	-20.0	0.97	-7.17	8.0	2.09	0.22	-31.40	5.11	9.57	5.93	-17.7
	Proteinous	29.14		35.39		2.89		0.22		143.88		173.40	
12	Frc.II	±	-18.7	±	43.26	±	0.67	±	-36.72	±	-2.81	±	-0.16
12	40mg/10 ml	0.71	10.7	1.10	13.20	3.9	0.07	0.13	30.72	5.12	2.01	13.84	0.10

<sup>\*</sup> Refers significance at P< 0.05 compared with control group. The values are mean  $\pm$  SE of 4 mice each group.

### References

- Kluwer W. "Facts and comparison: the review of natural products". St Louis, Mo., 1999;pp1-18.
- Whorter SM. "Biological complementary therapies: A focus on botanical product in diabetes". Diabetes spectrum. 2000;14:4.
- Rajasekeran S, Sivagnanam K, Subramanian. "Antioxidant effect of *Aloe vera* gel extract in streptozotocin-induced diabetes in rats". Phar Rep. 2005;57:90-96.
- Atherton P."Aloe vera revisited: Review of Aloe gel". Brit J Phytother. 1984; (4):176-183.
- Aguilar FJ, Ramos R, Saenz JL, Garcia FA. "Investigation on the hypoglycemic effects of extracts of four Mexican medicinal plants in normal and alloxan diabetic mice". Phyto Res. 2002;16:383-386.
- Gumieniczek A, Hopkala H, Wojtowich Z, Nikolajuk J. Changes in antioxidant status of heart musele tissue in experimental diabetes in rabbit. Acta Biochem Pol. 2002;49:529-535.

- Polidori MC, Stahl W, Eichlero N I, Sies H. Profiles of antioxidants in human plasma. Free Radic Biol Med. 2001;30:456-462.
- Larson RA. The antioxidants of higher plants. Phyto Chem.1988; 27: 969-978.
- Husain SM, Kasim MH., Cultivated plants of Iraq and their importance, Dar AL Kutub Organization for Printing and Publishing. University of Mosul, 1975.
- Bailely LH. Manual of cultivated plants, 16th Ed., Macmillan Publishing Co, Inc New York, 1977.
- Robyt JF, White BJ." Biochemical techniques theory and practice books cole publishing company". Monterey, California, 1987;391:268-269.
- Schacterle GR,Pollack PL. A simplified method for the quantitative assay of small amount of protein in biological material". Anal Biochem. 1973;51:654-655.
- 13. Andrews P. 1964; J.Biol.Chem., 96: 595.
- Alarcon. AFJ, Jimenez-E M, Roman-Ramos.RJ, "Hypoglycemic effects of extracts and fractions from psacalium decompositum in healthy and alloxan-diabetic mice". J Ethno Pharm. 2002;72:21-27.
- Neef H, Declercq P, Laekeman G. "Hypoglycemic activity of selected European plants". Phtother Res. 1995;9:45-48.
- Ahmad TY, AL-Khayat IK, Mahmood SZ. "Hypoglycemic activity of Olea europaea leaves". J Educ Sci. 1994;15:54-61.
- Tietz NW. Textbook of clinical chemistry,3rd Ed.C.A.Burtis, ER. Ashwood, WB.Saunders. 1999; pp826-835.
- Plummer D. An Introduction to practical biochemistry" 2nd Ed.Mc-Graw-Hill Book Co., 1978;pp.345-346.
- James RC, Goodman DR, Harbison RD. "Hepatic glutathione and hepatotoxicity, changes induced by selected marcortics". J Pharmaco Therap. 1982;221:708-714.

- Volken E, Nurperi G, Ahmet B. "N-acetyl cystine reduces cerebral lipid peroxidation in a rat model of infantile hydrocephalus". J Neurol Sci. 2001;Issue.1302-1310.
- Ahmed OA, "Modification and application of electrometric cholinesterase method of monitoring exposure to aganophosphate and carbamate insecticides". MSc Thesis, College of Pharmacy, University of Mosul, Mosul, Iraq 2001.
- Steel R, Torri J. "Principles and procedures of statistics". 2nd Ed.Mc.Graw-Hillbook, Co 1960; pp99-131.
- Muller T, Rocha J, Neis R. "Antidepessant inhibition human acetyl cholinesterase and butyrylcholinesterase activity". Biochem. Biophy Acta. 2002;1587:92-98.
- Gaurav KP. "Reversal of sodium nitrate induced airment of spontaneous alteration by *Aloe vera* gel. Involvement of cholinergic system", Pharmaco. Online. 2007;3:428-437.
- Talbott S, Hunghes K. "The health professional Guide to dietary supplements".http://supplement watch.com. 2006.
- Rahma A, Savsc A, Kumar V."Determination of total antioxidant activity of local vegetables shoots and the aqueous, ethanolic extracts gainst different concentration". Asia Pac J Clin Nutr. 203;12 (3):308-311
- Yeh G, Eisenberg D, Phillips R. Systematic review of herbs and dietary supplements for glycemic control in diabetes care. Eur J Clin Nutr. 2003;49:242-247.
- Nicotera P. Orrenius S. Role of thiols in protection against biological reactive intermediates. Adv Exp Med Biol. 1986;197: 41-49.
- Parihar M, Hemnani T. "Phenolic antioxidants attenuate hippocampal neuromal cell damage against kainic acid induced excitotoxicity". J
- 30. Biosci. 2003; 28 (1):121-128.
- Tirmenstein MA, Nicholls M. Glutathione depletion and the production of reactive oxygen species in isolated hepatocyte suspensions. Chem Biol. Interac. 2000;27:201-217.